# Upgrading Sterilization in the Orthodontics Practice: A Review

# Abstract

In today's orthodontic practice, disease control must undergo major reevaluation and restructuring. The knowledge of the natural history and treatment of many highly transmissible diseases to which orthodontic personnel are at high risk is changing rapidly. Recently a great deal of attention has been devoted to the spread of hepatitis b and aids viruses and the high risk of contamination of these viruses during the dental operations has made orthodontists more aware of the necessity of sterilization and disinfection. Finally, the most important ingredient to any change is in the orthodontic office staff that they must be enlightened, trained, and supervised by the orthodontist to effectively and efficiently switch from the old to the new.

#### **Key Words**

Cross-infection; instruments; orthodontic practice sterilization

Ezaz Ahmad<sup>1</sup>, Wamiq Shamim<sup>2</sup>, Saurabh Rathore<sup>3</sup>, Saurabh Bhatt<sup>4</sup>, Mohd Abdul Moiz<sup>5</sup>, Preeti Bhatt<sup>6</sup>

<sup>1</sup>Assistant Professor, Department of Orthodontics & Dentofacial Orthopaedics, Maharana Pratap College of Dentistry & Research Center, Gwalior, Madhya Pradesh, India

<sup>2</sup>Assistan Professor, Department of Orthodontics & Dentofacial Orthopaedics, Maharana Pratap College of Dentistry & Research Center, Gwalior, Madhya Pradesh, India

<sup>3</sup>Assistan Professor, Department of Orthodontics & Dentofacial Orthopaedics, Maharana Pratap College of Dentistry & Research Center, Gwalior, Madhya Pradesh, India

 <sup>4</sup>Assistan Professor, Department of Orthodontics & Dentofacial Orthopaedics, Maharana Pratap College of Dentistry & Research Center, Gwalior, Madhya Pradesh, India
<sup>5</sup>Consultant, Department of Orthodontics & Dentofacial Orthopaedics, Karnataka, India
<sup>6</sup>Assistant Professor, Department of

Prosthodontics, Hitkarni Dental College, Jabalpur, Madhya Pradesh, India

# INTRODUCTION

Sterilization is one of the most important methods in the dental office so that the orthodontic experience for the patient can be both fulfilling and completely safe. "Is the destruction of all microbial forms including viruses"<sup>[1-4]</sup>

# **PATHWAYS of Cross Contamination**

Routes and modes by which infection may spread in the dental clinic (Fig. 1).<sup>[5]</sup>

# Sterilization of Materials in Orthodontic Office

There are various method and sterilization machine used but most popularly used in dental office are Ultrasonic cleaning unit, Desktop autoclave sterilizer, Dry heat sterilizer, Ultraviolet cabinet , Chemical immersion or Cold sterilization, Glass bead sterilization.<sup>[1,2,6-8]</sup>

# Ultrasonic Cleaning Unit

Solutions with anti-rust compositions and enzyme based for breakdown of contaminants and particles is recommended. Pre-cleaning cycles last for 5-15 minutes (Fig. 2).

# **Desktop Autoclave Sterilizer**

Conventional method involves holding time of 15 minutes for  $121^{\circ}$  c, Rapid cycle involves  $134^{\circ}$  c for 3 minutes and cooling down period is of 40 minutes to 1 hour (Fig. 3).

# **Dry-Heat Sterilizer**

It causes oxidative destruction of bacterial protoplasm at a temperature range of  $160^{\circ}$  c ( $320^{\circ}$  F) for 2 hours to achieve complete sterilization and Rapid Dry - Heat sterilization involves cycles at temperatures of  $190^{\circ}$  c ( $375^{\circ}$  F) for 6 to 12 minutes (Fig. 4).

# **UV** Cabinet

Dental instrument and orthodontic pliers with 6



Sterilization in the orthodontics

81

Fig. 7

exposures of 5 minutes duration and the optimum wavelength for U V radiation is 260nm during peak emission. The range of 254 nm is suitable for adequate sterilization (Fig. 5).

# **Glass Bead Sterilization**

Sterilize the orthodontic bands and plier tips. They use small glass beads ranging from 1.2 to 1.5mm in diameter. The heating range is 217° c to 232° c for 3-5 seconds but not exceeding 250° c. Protocol involving molar band sterilization reported spore effectiveness at 226° c after 45 seconds for a single band (Fig. 6).<sup>[6]</sup>



Fig. 2



Fig. 4



Fig. 6



Fig. 8

Chemical immersion or cold sterilization

Suitable for heat-sensitive items, 2% acidic glutaralaldehyde and chlorine dioxide are commonly used. Most of the bacteria and virus are killed within 10 minutes. Sterilization time for instrument with 2% acidic glutaraldehyde is 10 hours without dilution and with chlorine dioxide is 6 hours.<sup>[13]</sup>

### **Orthodontic Plier Sterilization**

High quality stainless steel:- dry heat, autoclave, chemical vapor, ethylene oxide. Low quality stainless steel:- dry heat, chemical vapor, ethylene

#### **Blood Borne Pathogens And Other Disease Agents**

	•
MICROORGANISM	DISEASE
Hepatitis B (HBV)	Hepatitis B
Hepatitis C (HCV)	Non -a non-B, hepatitis
Hepatitis D (HDV)	Delta hepatitis
Herpes simplex type I	Oral herpes, Herpetic Whitlow ,Herpatic keratitis
Herpes simplex type II	Genital herpes
HIV	AIDS & ARC
Neisseria gonorrhoeae	Gonorrhoea
Treponema pallidum	Syphilis
Pseudomonas aeruginosa	Wound infections, abscesses
Staphylococcus aures	Wound infection, abscesses
Clostridium tetani	Tetanus
Micro Organism	s Transmitted By Inhalation
Micro organism	Disease
Varicella virus	Chicken pox
cytomegalovirus	Infections in infants
Measles (rubeola)	Measles and mumps
Influenza virus	Influenza and common cold
Rubella virus	German measles

Rubella vilus	German measies
Myobacterium tuberculosis	Tuberculosis
Sterptococcus pyogens	Oral abscesses, rheumatic fever & endocarditis

#### Candida albicans

oxide. Ultrasonic cycle for 5 to 12 minutes, rinsing with distilled water. Remove excess moisture thorough drying with compressed air (Oil-Free). Lubrication of Plier joints and cutting surfaces with silicone based lubricants. Sterilization protocol using a Dry-Heat Sterilizer at 190° c for 6 to 12 minutes with the placement of pliers in an open (Fig. 7a & Fig. 7b).<sup>[3,10]</sup>

# **Molar Bands**

- Ultrasonic cycle for 5 minutes depending on the capacity of the unit.
- Rinsing with distilled Water.
- Remove excess moisture through drying with Compressed Air (*Oil-Free*). .
- Sterilization protocol using a Dry-Heat Sterilizer at 190 ° c for 6 minutes.<sup>[12]</sup>

# STORAGE

# **Impression trays**

#### candidosis

- Aluminium or chrome plated: heat or gas sterilization
- Custom acrylic trays: disinfect by NaoCl or iodophor
- Plastic trays: ethylene oxide sterilization or disinfection by NaoCl or iodophor<sup>[13]</sup>

# **Dental Cast**

ADA recommends disinfection of stone cast by immersing in 1:10 NaoCl or an iodophor

#### **Orthodontic marking pencil**

Conventional orthodontic pencil can not be autoclave, gas sterilization can be used for alcohol containing permanent markers if possible always use disposable marker.

#### **Elastomeric Ligatures and Chains**

Single patient packs are the best insurance against cross-contamination risks at present and where this is not feasible as in the case of e-chain spools, it is better to cut a little extra than required and discard the rest.

# **Orthodontic Wire**

- TMA WIRE : Ethylene oxide ; autoclaving
- NiTi wire : Ethylene oxide
- STAINLESS STEEL WIRE : autoclave dry heat ; ethylene oxide<sup>[4,8]</sup>

# Alginate impression and Orthodontic Appliances

1% sodium hypochlorite, & 2% glutaraldehyde but not more than 10 minutes. Spraying aerosols are not recommended for their unevenness and additional inhalation risk.

#### **Floor Covering**

Should be impervious and non-slip . Carpeting must be avoided it should be Seam free and easy to clean.

# Disposal of clinical waste

Clinical waste sacks must be no more than three quarter full. Sharp waste (needle and scapel blades) must sealed in puncture proof container and disposed properly.

# INFECTION CONTROL PRACTISES FOR THE DENTAL OFFICE

### **Barrier Techniques**

Gloves

- Contact with body fluids, mucous membranes touching contaminated surfaces or items.
- After removal of gloves, wash hands, reglove.
- Repeated use by disinfection unacceptable.

# Heavy rubber

(Utility gloves) – Cleaning instruments and environmental surfaces. Allergy to latex or cornstarch – nylon glove linens. Polyethylene gloves – avoid contamination of objects

# **Protective Clothing**

Gowns, aprons, lab coats. Should not be warn outside work area.

# Masks

Syrgical marks – face, oral mucosa, nasal mucosa protection. Changed when soiled or wet.

#### **Protective Eyewears**

Eye protection from spatter of body fluids. OSHA Standard – Should have solid side shields.

# Limiting Contamination

- High volume evacuations
- Proper patient positioning > Avoid spatter
- Rubber dams.

Avoid contact with charts, telephones cabinets during treatment. Use  $2^{nd}$  pair of gloves or sheet of plastic wrap or fail.

# HANDS

Hand Washing

Ahmad E, Shamim W, Rathore S, Bhatt S, Moiz MA, Bhatt P

Washed at start of day, before gloving, after removal of gloves, after touching objects.

Routine procedures – soap and water. Surgical procedures – Antimicrobial hand scrub. Care must be taken when working with sharp object such as needle stick injury.

# HANDLING OF SHARP INSTRUMENTS AND NEEDLES

- Recap syringe needles.
- Disposable needles should not be bent or broken after use.
- Forceps used to handle sharp items.
- Discarded into puncture resistant biohazard containers.

# **OPERATORY SURFACES**

- Counter tops
- Light handles
- X ray unit heads
- Amalgamators
- Cabinet and drawer pulls
- Tray tables
- Chair switches
- Cover with plastic wrap, aluminium fact absorbent paper.
- Changed between patients.

# DISPOSAL OR WASTE MATERIALS

# Gloves, masks, wipes, paper drapes:

• Handled with gloves, discarded in impervious plastic bags.

#### Blood, disinfectants, sterilants

• Carefully poured into a drain connected to a sanitary server system.

#### Sharp items, needles, blades, scalpels

• Puncture- resistant containers marked with biohazard label.

#### Human tissue

• Same as sharp items, but diff. containers.<sup>[3,5,7,8]</sup>

# CONCLUSION

Elastomeric Ligatures and Chains are best single used. Current recommendations involve the immersion of alginate impressions and Orthodontic materials for not more than 10 minutes .The same infection control procedure must be used for all patients. Work surfaces should be impervious and easy to clean and disinfected .All autoclaves must be regularly serviced and maintained. Single use should be used whenever possible. All waste in the practice should be segregated. All staff understand the principle of personal protection.

# REFERENCES

1. Payne GS. Sterilization And Disinfection In The Orthodontic Office: A Practical Approach.

#### 84 Sterilization in the orthodontics

Am J Orthod Dentofacial Orthop. 1986;90(3):250-252.

- 2. Hamsa PRR. Instrument sterilization in the orthodontic clinic. JPSER. 2010;15-20.
- Vinay P, Reddy GY, Hegde N, Priyadarshini. Sterilization Methods in Orthodontics. International Journal of Dental Clinics. 2011:3(1):44-47.
- Sreekanth A, Cherukur I, Reddy M, Kumar L. A comparative evaluation of effect of different kinds of sterilizations on load defection characteristics, tensile properties of copper NITI wire. Annals and Essences of Dentistry. 2011;3(3):14-16.
- Shah R, Collins JM, Hodge TM, Laing ER. A national study of cross infection control: 'are we clean enough? British Dental Journal. 2009;6:207.
- Smith GE. Glass bead sterilization of orthodontic bands. Am J Orthod Dentofacial Orthop. 1986;90(3):243-249.
- Robert G. Savannah C. Trends in sterilization and disinfection procedures in orthodontic offices. Am J Orthod Dentofacial Orthop. 1990;98(4):292-299.
- Johnston, Moore, Rodu. Comparison of convection heat sterilization units. Am J Orthod Dentofacial Orthop. 1991:57-63.
- 9. Burke, Lewis, Wilson. Sterilization and disinfection special article. Am J Orthod Dentofacial Orthop. 1991:477-481.
- Payne, Masunaga, Alizadeh CM. Sterilization and disinfection. Am J Orthod Dentofacial Orthop. 1986:250-252.
- 11. Kirchhoff ST, Sekijima RK, Mark. JCO. 1987: 326-336.
- 12. Hohlt WF, Miller CH, Neeb JM, Sheidrake MA. Sterilization of orthodontic instruments and bands in cassettes. Am J Orthod Dentofacial Orthop. 1990:411-416.
- Hohlt, Miller, Neeb, Sheldrake. Sterilization of orthodontic instruments and bands in cassettes. Am J Orthod Dentofacial Orthop. 1988:117-122.